



# Data Sheet

## **Thiamphenicol (TAP)ELISA Kit**

**Cat. #ELAB39**

**Size: 96 Wells**



## Principle and Application

This kit adopts the method of indirect competitive enzyme-linked immunoassay (ELISA) to detect Thiamphenicol (TAP) in the sample such as livestock tissue, eggs, honey. The kit is composed of Microtiter Plate coated with coupled antigens, HRP enzyme conjugates, antibodies, standards and other supporting reagents. During the detection, with adding standards or samples, the TAP in the samples will compete with the coupled antigens to combine with anti-TAP antibodies. After adding enzyme conjugates, take coloration with TMB substrates. Absorbance value of the samples is a negative correlation with TAP content. Lastly, by comparing the obtained absorbance values with the standard curve, we can calculate the content of TAP in the sample.

## Storage conditions

- The kit shall be stored at 2-8 °C. Avoid freezing.
- Shelf Life: 12 months. The date of manufacture is presented in the label of the box.

## Technique Data

- Kit Sensitivity: 0. 5ppb (ng/mL)
- Reactive Mode: 25°C, 30min~15min
- Detection Limits:

Sample	Detection Limits
Tissue, honey	0.25ppb
Eggs	1ppb

- Cross-reaction Rate:

Thiamphenicol.....100%  
Florfenicol.....36.4%  
Chloramphenicol .....308%

- Sample Recovery Rate:

Sample	Recovery Rate
Tissue	85±20%
Honey	85±25%
Eggs	85±25%

## Composition of the Kit

Reagent	Specification
Microtiter Plate	8wells× 12strips
Standard: 0ppb, 0. 5ppb, 1.5ppb, 4.5ppb, 13.5ppb, 40.5ppb	1.0mL each
High Standard: 100ppb(red cap)	1×1.0mL
Antibody solution (blue cap)	1×5.5mL
HRP conjugate (red cap)	1×5.5mL
Substrate Reagent A (white cap)	1×6mL
Substrate Reagent B (black cap)	1×6mL
Stop Solution (yellow cap)	1×6mL
Concentrated Wash Buffer (20×)(white cap)	1×40mL
Concentrated Reconstitution Buffer (2×) (yellow cap)	1×50mL
Instruction	1
Adhesive Membrane	1
Sealed bag	1

## Materials Required but Not Supplied

- **Equipment:** microplate reader, printer, grinder (for homogenizing solid samples), nitrogen evaporator, vortex mixer (for shake and mix), centrifuge, graduated transfer pipette, and balance with a division value of 0.01 g, constant temperature device;
- **Micropipette:** single-channel 20 to 200µL and 100 to 1000µL, and multi-channel 300µL;

- **Reagents:** Ethyl acetate, n-hexane, acetonitrile.

## **Experimental preparation**

***Restore all reagents and samples to room temperature (adjust to around 25°C) for more than 30 min before use. This is a crucial step to ensure there is no precipitation in the reagents.***

Please note that the labware must be clean. Use disposable pipette tips to avoid contamination of interference results.

### **◆ Solution preparation:**

#### **Solution 1:** Reconstitution Buffer

Dilute the Concentrated Reconstitution Buffer (2×) 2 times with deionized water (Reconstitution Buffer (2×): deionized water=1:1). It can be stored at 4 °C for one month.

#### **Solution 2:** Working Wash Buffer

Dilute the concentrated wash buffer (20×) by a factor of 20 (Concentrated wash buffer/Deionized water= 1: 19).

#### **Solution 3:** Acetonitrile-Water Solution

Mix acetonitrile and water in a volume ratio of 84:16.

### **◆ Sample pretreatment steps:**

#### **1. Tissue treatment.**

- 1) Weigh 3g ± 0.05g of homogenized sample into a 50mL centrifuge tube, add 6mL of ethyl acetate, and shake for 5 minutes. Centrifuge at 4000 rpm at room temperature for 10 minutes.
- 2) Transfer 4mL of the upper layer to a container and evaporate to dryness under nitrogen or air at 50°C-60°C.

3) Dissolve the dried residue in 1mL of n-hexane, mix thoroughly, then add 1mL of **reconstitution buffer (Solution 1)**, shake for 2 minutes, and centrifuge at 4000 rpm at room temperature for 10 minutes

4) Discard the upper organic phase and take 50 $\mu$ L of the lower aqueous phase for analysis.

***Dilution times of the sample:0.5    Detection limits:0.25ppb***

## **2. Honey treatment.**

1) Weigh 2g  $\pm$  0.05g of honey into a centrifuge tube, dissolve it with 4mL of deionized water, then add 4mL of ethyl acetate, and shake for 2 minutes. Centrifuge at 4000 rpm at room temperature for 10 minute.

2) Transfer 2mL of the upper layer to a container and evaporate to dryness under nitrogen or air at 50°C-60°C.

3) Dissolve the dried residue in 0.5mL of **reconstitution buffer (Solution 1)**, and mix thoroughly.

4) Take 50 $\mu$ L for analysis.

***Dilution times of the sample:0.5    Detection limits: 0.25ppb***

## **3. Egg treatment.**

1) Weigh 3g  $\pm$  0.05g of homogenized egg sample into a 50ml centrifuge tube. Add 9mL of **acetonitrile-water solution (Solution 3)**, mix by shaking for 2 minutes, and centrifuge at 4000r/min at room temperature for 10 minutes.

2) Transfer 3mL of the upper layer liquid to another centrifuge tube, add 3mL of deionized water, and then add 4mL of ethyl acetate. Shake for 1 minute and centrifuge at 4000r/min at room temperature for 10 minutes.

3) Transfer 2mL of the upper layer liquid to a clean, dry glass tube and evaporate under nitrogen or air at 50°C-60°C until dry.

- 4) Dissolve the dried residue with 1ml of n-hexane, then add 1ml of **reconstitution solution (Solution 1)**. Mix for 30 seconds and centrifuge at 4000r/min at room temperature for 5 minutes.
- 5) Remove the upper organic phase, and take 50 $\mu$ L of the lower aqueous phase for analysis.

***Dilution times of the sample:2    Detection limits: 1ppb***

## **ELISA procedure**

Place all reagents and samples to room temperature (adjust to around 25°C) for 30min. Gently shake the reagent bottles before use.

Take out the frame of the microplate along with the required number of wells. Then place the unused microplate wells into the sealed bag with the desiccant provided. Store the remaining kit in the refrigerator at 2-8°C.

**Step 1: Number:** Number the wells in sequence corresponding to the samples and standard, make 2-well parallel trials for each sample and standard, and record their locations.

**Step 2: Incubation:** Add 50 $\mu$ L of **standard or sample** into each numbered well, then add 50 $\mu$ L of **HRP conjugate** per well. Next, add 50 $\mu$ L of **antibody solution** into each well. Finally, cover the Microtiter Plate with the adhesive membrane, shake gently by hand (or use a microplate shaker) for 5s and incubate for 30 min at 25°C in the dark.

**Step 3: Washing:** Uncover the adhesive membrane carefully, discard liquid in the wells, pipette 350 $\mu$ L of **Working Wash Buffer (Solution 2)** to every well, let stand for 30 seconds then drain, repeat 5 times. Invert the plate and tap it against a thick absorbent paper (or lint-free cloth), with a soft towel placed underneath. (Bubbles that are not removed after tapping dry can be punctured with a clean pipette tip).

**Step 4: Color:** Add 50 $\mu$ L of **Substrate Reagent A** to each well. Then add 50 $\mu$ L of **Substrate Reagent B** per well. Shake gently by hand (or use a microplate shaker) for 5s, and allow to react for 15min at 25°C in the dark. (The reaction can be extended

appropriately if the blue color is too pale.)

**Step 5: Stop the reaction:** Pipette 50µL of **Stop Solution** to each well, and shake gently by hand (or use a microplate shaker). The reaction would be stopped.

**Step 6: Calculate:** Determine the Optical Density (OD value; absorbance value) at 450nm (Reference wavelength 630nm) with a microplate reader. Finish this step within 10min after stop the reaction.

## Interpretation of result

### ◆ Calculate the percentage of absorbance value

$$\text{Percentage of absorbance value(\%)} = \frac{A}{A_0} \times 100\%$$

A—the average OD value of the sample or standard;

A<sub>0</sub>—the average OD value of the 0ppb standard.

It is used to calculate the percentage absorbance of a standard or sample.

### ◆ Draw the standard curve and calculate

- Take absorbance percentage (A/A<sub>0</sub>) of standards as Y-axis and the corresponding log of standards concentration (ppb) as X-axis.
- Draw the standard semi-log curves with X-axis and Y-axis.
- Take absorbance percentage of samples substitute into standard curve, then can get the corresponding concentration from standard curve. **Last, the resulting concentration values multiplied by the corresponding dilution times is the actual concentration of TAP of samples.**

If professional analysis software of the kit is used for calculation, it is more convenient for accurate and rapid analysis of a large number of samples.

## Attention

- Before test, the reagents and samples should be balanced to room temperature (25°C). If below 25°C, it will lead to all the standard OD value on the low side.

- In the washing process, dry wells may result in non-linear standard curves and undesirable reproducibility. Therefore, proceed to the next step immediately after washing.
- Please mix the contents within the wells uniformly and wash the plate thoroughly. The reproducibility is largely determined by consistency of washing step.
- During the incubation, cover microplates with adhesive membrane to avoid light.
- Do not use kits that are overdue. Do not mix reagents with those from other lots.
- Substrate Reagent A/B is colorless. If not, please discard.
- If absorbance value of 0ppb is below 0.5 ( $A_{450nm} < 0.5$ ), it means that the reagent may be metamorphic.
- Stop solution is corrosives, please avoid contact with skin.
- **As the OD values of the standard curve may vary according to the conditions of actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test.**
- **For the mentioned sample, fast and efficient extraction methods are included in the kit description. Please consult technical support for the applicability if other sample need to be tested.**
- The kit is used for rapid screening of actual samples. If the test result is positive, the instrument method such as HPLC, LC/MS can be used for quantitative confirmation.



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