



# Data Sheet

## **Ofloxacin (OFX) ELISA Kit**

**Cat. #ELAB40**

**Size: 96 Wells**



## Principle and Application

This kit adopts the method of indirect competitive enzyme-linked immunoassay (ELISA) to detect Ofloxacin (OFX) in tissue, honey, milk and other samples. The kit is composed of Microtiter Plate coated with coupled antigens, HRP conjugate, antibodies, standards and other supporting reagents. During the detection, with adding standards or samples, the OFX in the samples will compete with the coupled antigens to combine with anti-OFX antibodies. After adding HRP conjugate, take coloration with TMB substrates. Absorbance value of the samples is a negative correlation with OFX content. Lastly, by comparing the obtained absorbance values with the standard curve, we can calculate the OFX content in the sample.

## Storage conditions

- The kit shall be stored at 2-8 °C. Avoid freezing.
- Shelf life: 12 months. The date of manufacture is presented in the label of the box.

## Technique Data

- Kit Sensitivity: 1.5ppb (ng/mL)
- Reactive Mode: 25°C, 45min~15min
- Detection Limits:

Sample	Detection Limits
Tissue (Chicken, pork, fish, shrimp, etc.)	3ppb
Honey	3ppb
Milk	30ppb
Milk powder	75ppb
Poultry egg	45ppb
Urine	7.5ppb
Serum	45ppb

- Cross-reaction Rate:

Ofloxacin.....100%

- Sample Recovery Rate:

Sample	Recovery Rate
Tissue, honey, milk, milk powder, poultry eggs, urine, egg	85±15%

## Composition of the Kit

Reagent	Specification
Microtiter Plate	8wells× 12strips
Standard: 0ppb, 1.5ppb, 4.5ppb, 13.5ppb, 40.5ppb, 121.5ppb	1.0mL each
High Standard (red cap): 100ppb	1×1.0mL
Antibody solution (blue cap)	1×5.5mL
HRP conjugate (red cap)	1×5.5mL
Substrate Reagent A (white cap)	1×6mL
Substrate Reagent B (black cap)	1×6mL
Stop Solution (yellow cap)	1×6mL
Concentrated Wash Buffer (20×)(white cap)	1×40mL
Concentrated Reconstitution Buffer (5×)(yellow cap)	1×50mL
Instruction	1
Adhesive Membrane	1
Sealed bag	1

## Materials Required but Not Supplied

- **Equipment:** microplate reader, printer, grinder (for homogenizing solid samples), nitrogen evaporator, vortex mixer (**for shake and mix**), centrifuge, graduated transfer pipette, and balance with a division value of 0.01 g, constant temperature device;
- **Micropipette:** single-channel (20-200µL and 100-1000µL), and multi-channel 300µL;

- **Reagents:** Anhydrous Acetonitrile, n-Hexane, Concentrated Hydrochloric Acid, Dichloromethane.

## **Experimental preparation**

***Restore all reagents and samples to room temperature (adjust to around 25°C) for more than 30 min before use. This is a crucial step to ensure there is no precipitation in the reagents.***

Please note that the labware must be clean. Use disposable pipette tips to avoid contamination of interference results.

### **◆ Solution preparation:**

#### **Solution 1:** 0.15 M Hydrochloric Acid Solution

Dissolve 5 mL of concentrated hydrochloric acid in deionized water, and make up to 400mL.

#### **Solution 2:** Sample extraction solution

Take 10 mL of 0.15 M hydrochloric acid solution (Solution 1) and add it to 90 mL of anhydrous acetonitrile, mix thoroughly.

#### **Solution 3:** Reconstitution Buffer

Dilute the Concentrated Reconstitution Buffer (5×) five times (Concentrated Reconstitution Buffer (5×) /Deionized water= 1:4) .The Reconstitution Buffer can be stored for one month at 4°C.

#### **Solution 4:** Working Wash Buffer

Dilute the concentrated wash buffer (20×) by a factor of 20,  
(Concentrated wash buffer/Deionized water= 1: 19)

### **◆ Sample pretreatment steps:**

#### **1. Tissue (Chicken, pork, fish, shrimp, etc.) treatment.**

1) Transfer 2g±0.05g homogenized tissue sample to a 50mL centrifuge tube.

- 2) Add 8 mL of **Sample extraction solution (Solution 2)**. Vortex for 5 minutes and then centrifuge at room temperature at 4000 rpm for 10 minutes.
- 3) Transfer 2mL of the clear upper layer to a clean and dry 10mL glass test tube, and evaporate to dryness under nitrogen or air at 50°C-60°C.
- 4) Add 1 mL of **n-hexane**, shake for 2 minutes, then add 1 mL of **Reconstitution Buffer (Solution 3)**, shake for 30 seconds, centrifuge at 4000 rpm for 5 minutes at room temperature.
- 5) Remove the upper layer. Take 50µL of the lower layer for analysis.

***Dilution times of the sample:2      Detection limits: 3ppb***

## **2. Honey treatment.**

- 1) Transfer 1g±0.05g of honey to a 50ml centrifuge tube, add 6mL of **Sample extraction solution (Solution 2)**, shake for 5 minutes to ensure complete dissolution.
- 2) Add 3mL of **Reconstitution Buffer (Solution 3)**, add 11ml of **dichloromethane**, shake for 5 minutes, centrifuge at 4000 rpm at room temperature for 5 minutes.
- 3) Remove the upper layer, transfer the lower liquid (8 mL) to a dry container, and evaporate to dryness under nitrogen or air at 50°C-60°C.
- 4) Dissolve the dried residue in 1 mL of **Reconstitution Buffer (Solution 3)**, then add 1 mL of **n-hexane**, mix for 30 seconds, centrifuge at 4000 rpm at room temperature for 5 minutes.
- 5) Remove the upper layer. Take 50µL of the lower layer for analysis.

***Dilution times of the sample:2      Detection limits: 3ppb***

## **3. Milk treatment.**

- 1) Mix 25µL of the sample with 475µL of **Reconstitution Buffer (Solution 3)**, shake for 1 minute to ensure complete dissolution.
- 2) Take 50µL for analysis.

***Dilution times of the sample:20      Detection limits: 30ppb***

#### 4. Milk powder treatment.

- 1) Weigh  $0.5\text{g}\pm 0.05\text{g}$  of the sample into a 10mL centrifuge tube, add 5mL of deionized water, shake to ensure complete dissolution.
- 2) Mix 100 $\mu\text{L}$  of the sample solution with 400 $\mu\text{L}$  of **Reconstitution Buffer (Solution 3)**, shake for 1 minute.
- 3) Take 50 $\mu\text{L}$  for analysis.

***Dilution times of the sample:50    Detection limits: 75ppb***

#### 5. Poultry egg treatment.

- 1) Take  $1\text{g}\pm 0.05\text{g}$  of homogenized egg liquid into a 10mL centrifuge tube, add 5mL of deionized water, shake to ensure thorough mixing.
- 2) Mix 100 $\mu\text{L}$  of the sample solution with 400 $\mu\text{L}$  of **Reconstitution Buffer (Solution 3)**, shake for 1 minute.
- 3) Take 50 $\mu\text{L}$  for analysis.

***Dilution times of the sample:30    Detection limits: 45ppb***

#### 6. Urine treatment

- 1) Mix 4mL of **reconstitution solution (Solution 3)** with 1mL of centrifuged clear urine, and shake for 30 seconds.
- 2) Take 50 $\mu\text{L}$  of the liquid for analysis.

***Dilution times of the sample: 5    Detection limit: 7.5ppb***

#### 7. Serum treatment

- 1) Mix 50 $\mu\text{L}$  of serum sample with 1450 $\mu\text{L}$  of **reconstitution buffer (Solution 3)**, and shake for 1 minute.
- 2) Take 50 $\mu\text{L}$  of the liquid for analysis.

***Dilution times of the sample:30    Detection limit: 45ppb***

## ELISA procedure

Place all reagents and samples to room temperature (adjust to around 25°C) for 30min.

Gently shake the reagent bottles before use.

Take out the frame of the microplate along with the required number of wells. Then place the unused microplate wells into the sealed bag with the desiccant provided. Store the remaining kit in the refrigerator at 2-8°C.

**Step 1: Number:** Number the wells in sequence corresponding to the samples and standard, make 2-well parallel trials for each sample and standard, and record their locations.

**Step 2: Incubation:** Add 50µL of **standard or sample** into each numbered well, then add 50µL of **HRP conjugate** per well. Next, add 50µL of **antibody solution** into each well. Finally, cover the Microtiter Plate with the adhesive membrane, shake gently by hand (or use a microplate shaker) for 5s and incubate for 45 min at 25°C in the dark.

**Step 3: Washing:** Uncover the adhesive membrane carefully, discard liquid in the wells, pipette 350µL of **Working Wash Buffer (Solution 4)** to every well, let stand for 30 seconds then drain, repeat 5 times. Invert the plate and tap it against a thick absorbent paper (or lint-free cloth), with a soft towel placed underneath. (Bubbles that are not removed after tapping dry can be punctured with a clean pipette tip).

**Step 4: Color:** Add 50µL of **Substrate Reagent A** to each well. Then add 50µL of **Substrate Reagent B** per well. Shake gently by hand (or use a microplate shaker) for 5s, and allow to react for 15min at 25°C in the dark. (The reaction can be extended appropriately if the blue color is too pale.)

**Step 5: Stop the reaction:** Pipette 50µL of **Stop Solution** to each well, and shake gently by hand (or use a microplate shaker). The reaction would be stopped.

**Step 6: Calculate:** Determine the Optical Density (OD value; absorbance value) at 450nm (Reference wavelength 630nm) with a microplate reader. Finish this step within 10min after stop the reaction.

## Interpretation of result

### ◆ Calculate the percentage of absorbance value

$$\text{Percentage of absorbance value(\%)} = \frac{A}{A_0} \times 100\%$$

A—the average OD value of the sample or the standard;

A<sub>0</sub>—the average OD value of the 0ppb standard.

It is used to calculate the percentage absorbance of a standard or sample.

### ◆ Draw the standard curve and calculate

- Take absorbance percentage(A/A<sub>0</sub>) of standards as Y-axis and the corresponding log of standards concentration (ppb) as X-axis.
- Draw the standard semi-log curves with X-axis and Y-axis.
- Take absorbance percentage of samples substitute into standard curve, then can get the corresponding concentration from standard curve. **Last, the resulting concentration values multiplied by the corresponding dilution times is the actual concentration of OFX of samples.**

If professional analysis software of the kit is used for calculation, it is more convenient for accurate and rapid analysis of a large number of samples.

## Attention

- Before test, the reagents and samples should be balanced to room temperature (25°C). If below 25°C, it will lead to all the standard OD value on the low side.
- In washing process, dry wells may result in non-linear standard curves and undesirable reproducibility. Therefore, proceed to the next step immediately after washing.
- Please mix the contents within the wells uniformly and wash the plate thoroughly. The reproducibility is largely determined by consistency of washing step.
- During the incubation, cover microplates with adhesive membrane to avoid light.

- Do not use kits that are overdue. Do not mix reagents with those from other lots.
- Substrate Reagent A/B is colorless. If not, please discard.
- If absorbance value of 0ppb is below 0.5 (A450nm < 0.5 ), it means that the reagent may be metamorphic.
- Stop solution is corrosives. Please avoid contacting with skin.
- **As the OD values of the standard curve may vary according to the conditions of actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test.**
- **For the mentioned sample, fast and efficient extraction methods are included in the kit description. Please consult technical support for the applicability if other sample need to be tested.**
- The kit is used for rapid screening of actual samples. If the test result is positive, the instrument method such as HPLC, LC/MS can be used for quantitative confirmation.



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